

## **FOOD MICROBIOLOGY - CYCLE 28**

### **NOTE: SEE NEW INSTRUCTIONS FOR USE**

#### **Introduction:**

It is important to read and understand this document. If you have any queries please contact Thistle QA immediately for assistance.

#### **Important Information:**

These devices contain viable organisms that may, under certain circumstances, produce disease. Proper techniques must be employed to avoid exposure and contact with any organism growth.

The laboratory must be equipped, and have the facilities to receive, process, maintain, store and dispose of biohazard material.

Personnel using these devices must be trained, experienced and demonstrate proficiency in processing, maintaining, storing and disposing of biohazard material.

Please handle the samples as routine samples.

**We request that you return results on all the samples, even if they do not grow.**

In the case of any discrepancy regarding the specimen your lab has received, please contact Thistle QA immediately so that an investigation can be done or the specimen number can be recalled for assessment by appropriate experts / supplier.

**Please check your kit upon arrival and call Thistle immediately if there are any problems with your kit.**

#### **Characteristics:**

The samples are packaged in re-sealable vials that contain pellets of microorganisms and desiccators to prevent adverse accumulations of moisture.

#### **Storage:**

Store the Lyophilised Microorganisms at 2 – 8°C in the original, sealed vial.

**The Lyophilised microorganism should not be used if:**

Stored improperly

There is evidence of excessive exposure to heat or moisture

The expiration date has passed

## **NEW INSTRUCTIONS FOR USE**

1. Remove the vial of the lyophilized pellet from refrigerated storage (2°C to 8°C) and allow the unopened vial to equilibrate to room temperature (22°C to 25°C).
2. **Dilution 1:** With a sterile forceps, remove the pellet and place into **10ml of pre-warmed** dilution fluid as stated in the laboratory SOP. It is **ESSENTIAL** that the dilution fluid **be PREWARMED to 35°C to 37°C**.
3. **Incubate** the inoculated dilution fluid at 35°C to 37°C for **thirty (30) minutes ONLY**. Following the incubation, **mix (preferably vortex)** the inoculated dilution fluid thoroughly.
4. **Dilution 2:** Take **1ml of dilution 1** and add to **9ml of diluent**.
5. **Dilution 3: Take 1ml of dilution 2 and add to 9ml of diluent.**
6. Use **1ml of dilution 1** and proceed with quantification method\*\* as stated in the laboratory protocol (Pour Plate or Spread Plated method). **Run samples in duplicate.**
7. Use **1ml of dilution 2** and proceed with quantification method\*\*\* as stated in the laboratory protocol (Pour Plate or Spread Plated method). **Run samples in duplicate.**
8. **Use 1ml of dilution 3 and proceed with quantification method\*\*\* as stated in the laboratory protocol (Pour Plate or Spread Plated method). Run samples in duplicate.**
9. Proceed with the complete quantitative testing procedure as set forth in the laboratory SOP.
10. Upon completion of the procedure, record the average test results for dilution **1, 2 and 3.**

**Please Note: Dilution 1 is a 10x dilution**

**Dilution 2 is a 100x dilution.**

**Dilution 3 is a 1000x dilution**

**The final results should be recorded as CFU's per pellet**

**\*\* For usable statistical results please use a non-selective medium such as Plate Count Agar, Tryptic Soy Agar or Nutrient agar.**

**\*\*\* To compare results with selective agars (agars used for specific pathogens) please specify media used and run both dilutions simultaneously (Selective and Non-Selective agars).**

**\*\*NB\*\* Please make sure that dilutions are thoroughly mixed before plating out.**

You will receive vials with the relevant organisms as ordered by your laboratory for your specific needs. With each organism you need to report the following information to Thistle:

- The vial number
- The organism name as given to you by Thistle on the "Organism Names sheet".
- Indicate if this organism is Present or Absent
- State the specific count in CFU per pellet for dilution 1, 2 and 3
- State the media, temperature and incubation time for each dilution.

**Safety:**

These products are for in-vitro use only.

These devices, and subsequent growth of these micro-organisms on culture media, are considered to be biohazard material. These devices contain viable organisms that may, under certain circumstances, produce disease. Proper techniques must be employed to avoid exposure and contact with any organism growth. The microbiology lab must be equipped, and have the facilities to receive, process, maintain, store and dispose of biohazard material. Personnel using these devices must be trained, experienced and demonstrate proficiency in processing, maintaining, storing and disposing of biohazard material.

Please fax or email the results back to us.

Reports will be e-mailed within 7 – 10 working days of the final cut-off date.

Collusion and/or falsification of EQA results are not good accreditation practice.

**Return of Results:**

Each of the vials has a number printed on the label. We recommend analysis dates as shown below. Please send your results - **by the latest** - on the final cut-off dates given below. If the recommended analysis date does not allow you to get results to us on time, please analyse earlier. Use the correct dates for the vial numbers in your laboratory's kit. If in doubt, please contact Thistle immediately for assistance.

<u>MONTH NUMBER:</u>	<u>VIAL NUMBER:</u>	<u>ANALYSIS DATES:</u>			<u>FINAL CUT-OFF DATES:</u>		
Month 1	1	24	May	2010	31	May	2010
	2	24	May	2010	31	May	2010
	3	24	May	2010	31	May	2010
Month 2	4	21	June	2010	28	June	2010
	5	21	June	2010	28	June	2010
Month 3	6	19	July	2010	26	July	2010
	7	19	July	2010	26	July	2010
	8	19	July	2010	26	July	2010
Month 4	9	23	August	2010	30	August	2010
	10	23	August	2010	30	August	2010
Month 5	11	20	September	2010	27	September	2010
	12	20	September	2010	27	September	2010
	13	20	September	2010	27	September	2010
Month 6	14	18	October	2010	25	October	2010
	15	18	October	2010	25	October	2010
	16	18	October	2010	25	October	2010